
Hypoxia, Angiogenesis and Atherogenesis

L. Heikal and G.A.A. Ferns

Additional information is available at the end of the chapter

Author Queries	
[AQ01]	Please provide full name of initials of both the authors.
[AQ02]	Please shorten the Abstract text to a maximum of 200 words.
[AQ03]	Ref. [85] is not provided in the reference list. Please check.
[AQ04]	Please provide page number in Ref. [2].
[AQ05]	Please provide the volume number in Refs. [65] and [67].
[AQ06]	Please provide the volume and page numbers in Ref. [83].

04

Abstract

05

06

07

08

09

10

11

12

13

14

15

16

17

18

19

20

21

22

23

The balance between vascular oxygen supply and metabolic demand for oxygen within the vasculature is normally tightly regulated. An imbalance leads to hypoxia and a consequential cascade of cellular signals that attempt to offset the effects of hypoxia. Hypoxia is invariably associated with atherosclerosis, wound repair, inflammation and vascular disease. The anoxaemia hypothesis proposes that an imbalance between the demand for and supply of oxygen in the arterial wall is a key factor in the development of atherosclerosis and plaque angiogenesis. There is now substantial evidence that hypoxia plays an essential role in angiogenesis as well as plaque angiogenesis. It controls the metabolism, and responses of many of the cell types found within the developing plaque and whether the plaque will evolve into a stable or unstable phenotype. Hypoxia is characterized in molecular terms by the stabilization of hypoxia-inducible factor (HIF) 1 α , a subunit of the heterodimeric nuclear transcriptional factor HIF-1 and a master regulator of oxygen homeostasis. The expression of HIF-1 is localized to perivascular tissues, inflammatory macrophages and smooth muscle cells where it regulates several genes that are important to vascular function including vascular endothelial growth factor, nitric oxide synthase, endothelin-1 and erythropoietin. This chapter summarizes the effects of hypoxia on the functions of cells involved in angiogenesis as well as atherogenesis (plaque angiogenesis) and the evidence for its potential importance from experimental models and clinical studies.

AQ02

24

Keywords: hypoxia, HIF-1, proliferation, atherosclerosis, plaque formation, blood vessel

25

26

1. Introduction

27

28

29

The circulatory system develops early in mammalian embryogenesis. An oxygen supply is essential for normal tissue function, development and homeostasis. The vascular network within the cardiovascular system is essential for the delivery of oxygen, nutrients and other

01 molecules to the tissues of the body [1]. Oxygen availability serves as an important regulator
02 of the cardiovascular system. Oxygen balance may be perturbed if there is reduced oxygen
03 diffusion, or increased oxygen consumption that may be a consequence of rapid cellular divi-
04 sion during embryonic development, by tumour growth, or by vasculature dysfunction due
05 to vessel occlusion or rupture [2].

06 Hypoxia is defined by a reduced oxygen tension relative to those normally extant within a
07 particular tissue. It has multiple impacts on the vascular system and cell function [3]. The
08 effects of moderate hypoxia (3–5% O₂) are usually reversible and are usually accompanied by
09 adaptive physiological responses in the cells. A lower oxygen tension (0–1% O₂) contributes
10 to the pathophysiology of tumour progression and cell apoptosis [4] and is a feature of condi-
11 tions that include cancer, ischemic heart disease, peripheral artery disease, wound healing
12 and neovascular retinopathy. Hypoxia promotes vessel growth by stimulating an upregula-
13 tion of multiple proangiogenic pathways that mediate key aspects of endothelial, stromal
14 and vascular support cell biology. The role of hypoxia in human disease is now becoming
15 increasingly clear [5] including the association between hypoxia and endothelial dysfunction
16 that affects several cellular processes and signal transduction.

17 Hypoxia can occur in several ways: (1) hypoxic hypoxia is caused by an insufficient oxygen
18 concentration in the air in the lungs, which may occur during sleep apnea, when the diffusion
19 of oxygen to the blood is reduced, or at high altitude; (2) hypoxemic hypoxia occurs when the
20 blood has reduced transport capacity as seen in carbon monoxide poisoning when haemo-
21 globin cannot carry oxygen at its normal concentrations; (3) stagnant hypoxia results when
22 the cardiac output does not match the demands of the body and the flow is not sufficient to
23 deliver enough oxygenated blood to the tissue and (4) histotoxic hypoxia occurs when cells
24 cannot utilize the available oxygen, for example following cyanide poisoning when oxygen
25 cannot be used to produce ATP as the mitochondrial electron transport is inhibited.

26 Chronic tissue hypoxia (an oxygen tension of 2–3% for a prolonged period of time) may cause
27 uncontrolled proliferation of cells. When physiological oxygen concentrations are restored,
28 the increased blood flow supplies excessive oxygen; this may then lead to increased free-
29 radical generation, tissue damage and concomitant activation of stress-response genes; a con-
30 dition known as ‘reoxygenation injury’. In these circumstances, normal cells/tissues may not
31 survive; but tumour cells are still able to proliferate despite the hypoxic milieu, as they have
32 developed genetic and adaptive changes leading to resistance to hypoxia [6].

33 Hypoxia also plays important roles in normal human physiology and development; for exam-
34 ple, it is integral to normal embryonic development. Whatever the cause, or the severity of
35 hypoxia, it leads to an induction of adaptive responses within the endothelial and vascular
36 smooth muscle cells through the activation of genes that participate in angiogenesis, cell pro-
37 liferation/survival and in glucose and iron metabolism [7].

38 In healthy vascular tissue, vascular smooth muscle cells (SMCs) and endothelial cells (ECs)
39 proliferate at very low levels. However, SMCs and ECs can be stimulated to re-enter the cell
40 cycle in response to several physiological and pathological stimuli. Hypoxia is considered an
41 important stimulus of SMC and EC proliferation and is found in atherosclerotic lesions and
42 rapidly growing tumours [4].

01 The proliferation of ECs is pivotal to the formation of new micro-vessels and is important
02 during organ development in embryogenesis and tumour growth, and also contributes to
03 diabetic retinopathy, psoriasis, rheumatoid arthritis and atherosclerosis. Abnormal SMC pro-
04 liferation contributes to atherosclerosis, intimal hyperplasia after angioplasty and graft ath-
05 erosclerosis after coronary transplantation [8, 9].

06 **2. Consequences of hypoxia**

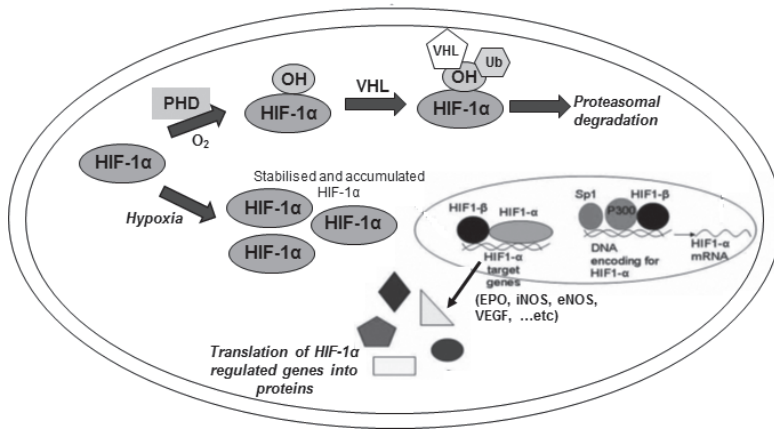
07 Most cells are able to survive under hypoxic conditions through the transcriptional activa-
08 tion of a series of genes. The oxygen-sensitive transcriptional activator, hypoxia-inducible
09 factor-1 (HIF-1) is the key transcriptional mediator of the hypoxic response and master
10 regulator of O₂ homeostasis. It orchestrates the profound changes in cellular transcription
11 that accompanies hypoxia by controlling the expression of numerous angiogenic, meta-
12 bolic and cell cycle genes. Accordingly, the HIF pathway is currently viewed as a master
13 regulator of angiogenesis [5].

14 HIF-1 is normally only found in hypoxic cells. It is a heterodimer that is composed of an
15 O₂-regulated HIF-1 α subunit and a constitutively expressed HIF-1 β subunit [10]. In the
16 α -subunit, there is an oxygen-dependent degradation (ODD) domain, where the 4-hydroxy-
17 proline formation is catalysed by proline-hydroxylase-2 (PHD-2). This leads to its ubiqui-
18 tination by the von Hippel-Lindau E3 ubiquitin ligase (VHL) and subsequent proteasomal
19 degradation under normoxic cellular conditions. This prevents the formation of a functional
20 HIF dimer [11]. Since PHDs require oxygen for their catalytic activity, and function as cellular
21 oxygen sensors, HIF degradation only occurs under normoxic conditions. Factor inhibiting
22 HIF-1 (FIH) protein, which hydroxylates HIF-1, also contributes to HIF-1 inactivation in nor-
23 moxic conditions, and thereby prevents the interaction of this subunit with the two transcrip-
24 tional co-activators of HIF-1: p300 and CREB-binding protein (CBP) which are essential for
25 HIF-1 transcription. Expression and stabilization of the HIF-1 complex is regulated through
26 feedback inhibition, as PHD-2 itself is activated by HIF-1 [12].

27 Under hypoxic conditions, HIF-1 protein is stable and active as a hydroxylase, VHL proteins,
28 and FIH are all inhibited by a lack of oxygen. HIF-1 is then able to interact with its co-acti-
29 vators and can dimerize with its constitutively expressed β -subunit [12]. Once stabilized, the
30 HIF-1 protein can bind to the regulatory regions of its target genes, inducing their expression;
31 these target genes include VEGF (vascular endothelial growth factor) [13], erythropoietin [14]
32 and nitric oxide synthase (NOS) [15, 16] and other proangiogenic factors such as PlGF (pla-
33 cental growth factor), or angiopoietins [12] (**Figure 1**).

34 It has been proposed that the induction of a pseudo-hypoxic response by inhibiting HIF pro-
35 pyl 4-hydroxylases may provide a novel therapeutic target in the treatment of hypoxia-asso-
36 ciated diseases [17].

37 Several small molecules, such as dimethylxalyl glycine [18], Roxadustat (FG-4592) [19] and
38 ZYAN1 [20], have been developed to inhibit prolyl hydroxylase domain-containing (PHD)
39 enzymes, and cause HIF activation [21]. These agents have been applied to the treatment of



01 **Figure 1.** Regulation of the hypoxia-inducible transcription factor (HIF-1 α) pathway. Under normal oxygen tensions (normoxia), prolyl hydroxylase (PHD) enzymes, von Hippel-Lindau protein (pVHL), the ubiquitin ligase complex (Ub) and factor inhibiting HIF-1 (FIH) are active leading to HIF-1 α proteasomal degradation. Under hypoxic conditions, PHD, pVHL and Ub are not active leading to its cytoplasmic accumulation of HIF-1 α . The HIF-1 α gene is transcribed in the nucleus with the help of specificity protein (Sp) 1, P300, and HIF-1 β leading to transcription of HIF target genes such as EPO, NOS and VEGF (adapted from references and [65]).

02 renal anaemia in which there is a deficiency of erythropoietin [22, 23]. The administration of
 03 these compounds is associated with an improved iron profile and an increase of endogenous
 04 erythropoietin production to near the physiological range. The clinical trials currently under-
 05 way aim to address whether PHD enzyme inhibitors will improve clinical end-points, includ-
 06 ing cardiovascular events [24]. PHD inhibitors have been reported to reduce blood pressure
 07 [22] and plasma cholesterol concentrations [19]. Hence, there is a good reason to believe that
 08 some PHD inhibitors will reduce cardiovascular endpoints in patients with renal disease.
 09 Whether they will benefit a broader category of patients with high risk of cardiovascular
 10 disease is difficult to predict.

11 Hydroxylase activity can be also rescued by mutating specific regions, or by adding cobalt
 12 ions to the cell, the latter of which presumably compete for iron-binding sites. Some hydroxy-
 13 lases in the prolyl family can be selectively inhibited by Adriamycin *in vitro*. Cobalt (II) and
 14 nickel (II) ions increase HIF-1 activity in cells, presumably because these ions displace iron
 15 from the active sites of 2-oxo-glutarate (2OG) hydroxylases [12].

16 It has been shown that HIF-1 α can be regulated by non-hypoxic stimuli such as lipopolysac-
 17 charides (LPS), thrombin and angiotensin II (Ang II) [25]. Hormones such as angiotensin II
 18 and platelet-derived growth factor stimulate the HIF pathway by increasing HIF-1 α protein
 19 levels through production of reactive oxygen species (ROS) within the cell. Although the exact
 20 mechanism for this is unclear, it appears to be entirely distinct from the hypoxia pathways.

01 Thrombin and other growth factors appear to increase angiogenesis through HIF-1 α protein
02 agonist mechanisms. Insulin similarly activates HIF-1 α through the action of multiple protein
03 kinases necessary for expression and function. p53 is responsible for promoting ubiquitina-
04 tion of HIF-1 α , and may be another possible target for enhancing HIF-1. Homozygous
05 deletion of the p53 gene has been found to cause HIF-1 activation [26]. Gene therapy may
06 eventually be used to increase HIF-1 levels and relieve complications of ischemia. For exam-
07 ple, delivery of a stabilized, recombinant form of HIF-1 α through adeno-associated virus
08 (AAV) in order to overexpress HIF-1 has been shown to result in significantly increased capil-
09 lary density in skeletal muscle [27]. While gene therapy approaches aimed at the process and
10 effects of angiogenesis continue to be developed and studied, higher levels of success in pre-
11 clinical trials currently are being sought before clinical applications are pursued. Amongst the
12 remaining obstacles in using gene therapy for this purpose is the effective mode of delivery
13 [12]. Inhibition of PHD2 using siRNA has been shown to decrease cardiac infarction size in
14 murine models [28, 29].

15 In addition to HIF-1 α , there are two other members of HIF superfamily that have been
16 described: HIF-2 and HIF-3 [30]. Both are important regulators of the hypoxia response
17 with similar actions as HIF-1 [31] and lead to the transcriptional activation of target
18 genes in hypoxia [32]. However, Eubank et al demonstrated opposing roles for the HIFs
19 in tumour angiogenesis, with HIF-1 exhibiting proangiogenic properties that act through
20 its effects on VEGF secretion, whereas HIF-2 exhibits anti-angiogenic activity by induc-
21 ing the production of the endogenous angiogenesis inhibitor, sVEGFR-1 [33]. HIF-3 α has
22 complementary functions, rather than redundant to HIF-1 α induction in protection against
23 hypoxic damage in alveolar epithelial cells in protection against hypoxic damage in alveo-
24 lar epithelial cells [34].

25 Although the oxygen-sensing mechanism involving oxygen-dependent hydroxylation of the
26 HIF- α subunits is probably a universal mechanism in cells, and has been highly conserved
27 during evolution, additional regulatory steps appear to determine which of the alternative
28 subunits is induced [34]. One of the best studied hypoxic responses that will be discussed in
29 this chapter is the induction of angiogenic factors and growth factors, which lead to the for-
30 mation and growth of new blood vessels.

31 **3. Hypoxia and angiogenesis**

32 Blood vessels formation occurs through two basic mechanisms: (1) vasculogenesis repre-
33 sents de novo formation of blood vessels, and is derived from endothelial progenitors and (2)
34 angiogenesis and arteriogenesis (formation of blood vessels from pre-existing blood vessels).

35 Angiogenesis is a tightly regulated multi-step process that begins when cells within a tissue
36 respond to hypoxia. When tissues grow beyond the physiological oxygen diffusion limit, the
37 relative hypoxia triggers expansion of vascular beds by inducing angiogenic factors in the
38 cells of the vascular beds, which are physiologically oxygenated by simple diffusion of oxy-
39 gen. Angiogenesis may be a physiological process, as in the case in embryonic development,

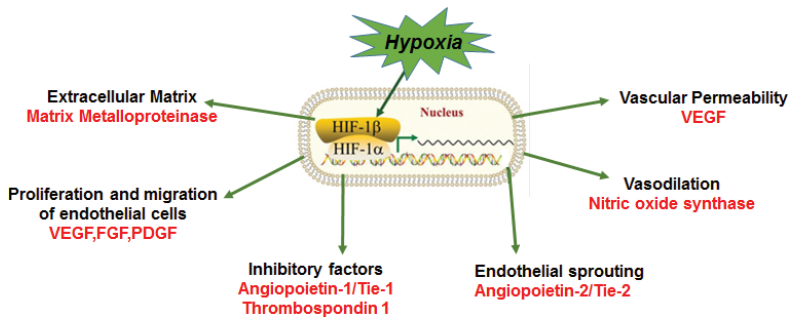
01 wound healing or vessel penetration into avascular regions. It may also be pathological, for
02 example when it occurs during the formation of solid tumours, eye disease, chronic inflamma-
03 tory disorders such as rheumatoid arthritis, psoriasis and periodontitis and atherosclerosis.

04 The regulation of angiogenesis (whether in physiological or pathological cases) by hypoxia
05 is an important component of homeostatic mechanisms that link vascular oxygen supply to
06 metabolic demand. An understanding of the processes involved in angiogenic, the role of
07 the interacting proteins involved, and how all this is regulated by hypoxia through an ever-
08 expanding number of pathways in multiple cell types may lead to the identification of novel
09 therapies and modalities for ischemic vascular diseases as well as diseases characterized by
10 excessive angiogenesis, such as rheumatoid arthritis, psoriasis, tumours, ischemic brain and
11 heart attack [5, 6].

12 Angiogenesis in hypoxia is regulated by several pro- and anti-angiogenic factors [1]. HIF-1
13 has been established as the major inducer of angiogenesis [35]. It regulates the transcription of
14 VEGF, a major regulator of angiogenesis which promotes endothelial cell migration towards
15 the hypoxic area. During hypoxia, HIF-1 binds to the regulatory region of the VEGF gene,
16 inducing its transcription and initiating its expression. VEGF is then secreted and binds to
17 cognate receptor tyrosine kinases (VEGFR1 and VEGFR2) located on the surface of vascular
18 endothelial cells triggering a cascade of intracellular signalling pathways that initiate angio-
19 genesis [10]. These endothelial cells are recruited to form new blood vessels which ultimately
20 supply the given area with oxygenated blood [12]. Interestingly, recent studies have shown
21 that hypoxia influences additional aspects of angiogenesis, including vessel patterning, matu-
22 ration and function [5].

23 Other factors such as angiopoietin-2/angiopoietin-1 [36, 37], angiopoietin receptor (Tie2) [38],
24 platelet-derived growth factor (PDGF) [39], basic fibroblast growth factor (bFGF) [40] and
25 monocyte chemoattractant protein 1 (MCP-1) [41] have also been reported to be responsible
26 not only for increasing vascular permeability, endothelial sprouting, maintenance, differen-
27 tiation and remodelling but also cell proliferation, migration, enhancement of endothelial
28 assembly and lumen formation (**Figure 2**). In hypoxia, angiogenesis is also modulated by
29 several factors that are secreted by leucocytes, which produce a high abundance of angiogenic
30 factors, various interleukins such as TGF- β 1 and MCP-1 and proteinases [42]. Thus, hypoxia
31 provides an important environmental stimulus not only for angiogenesis but also for related
32 phenomena in the hypoxic or surrounding area, suggesting that hypoxia is more than simply
33 a regulator of angiogenesis [6].

34 Angiogenesis may be detrimental when it is excessive. Therefore, angiogenic factors must be
35 highly active but also be tightly regulated. Angiogenesis that is associated with pathological
36 consequences may exhibit differences in the responsible molecular pathways in comparison
37 to physiological angiogenesis. Mutations in oncogenes and tumour suppressor genes and dis-
38 ruptions in growth factor activity play an important role in tumour angiogenesis. The activa-
39 tion of the most prominent proangiogenic factor VEGF might be due to physiological stimuli
40 such as hypoxia or inflammation or due to oncogene activation and tumour suppression func-
41 tion loss. Physiological angiogenesis that occurs during embryonic development or wound
42 healing seems to be dependent on VEGF signalling, whereas tumour angiogenesis adopts



01 **Figure 2.** HIF-1 α regulates factors involved in developmental and pathological angiogenesis. HIF-1 α directly regulates genes involved in steps such as vasodilation, increased vascular permeability, extracellular matrix remodelling and proliferation.

02 the ability to shift its dependence from VEGF to other proangiogenic pathways, for example,
 03 through the recruitment of myeloid cells and the upregulation of alternative vascular growth
 04 factors (PlGF and FGF) [1].

05 The identification of alternative ways of inhibiting tumour growth by disrupting the growth-
 06 triggering mechanisms of increasing vascular supply through angiogenesis will depend on
 07 the understanding of how tumour cells develop their own vasculature. Other cofactors are
 08 essential to ensure maximum efficiency of the transcriptional machinery related to changes
 09 in oxygen availability within cells/tissues, and the roles of different HIFs in eliciting hypoxic
 10 responses seem to be more divergent as originally assumed. Chen et al. have shown new
 11 regulatory interactions of HIF-related mechanisms involving the interactions of basic HIFs,
 12 HIF-1 α and HIF-2 α with their regulatory binding proteins, histone deacetylase 7 (HDAC7)
 13 and translation initiation factor 6 (Int6), respectively [6]. Int6 induces HIF-2 degradation. In
 14 addition, silencing of *Int6* produces a potent, physiological induction of angiogenesis that
 15 may be useful in the treatment of diseases related to insufficient blood supply. The newly
 16 discovered binding proteins-HDAC7 for HIF-1 and Int6 for HIF-2 support the assumption
 17 that the 2 HIF isoforms play distinct roles in eliciting hypoxia-related responses. HIF-2 may
 18 be considered as one of the master switches for inducing angiogenic factors at least in some
 19 cell types [6].

20 The hypoxia/reoxygenation cycle leads to the formation of reactive oxygen species (ROS)
 21 that may subsequently regulate HIF-1 but in a rather complex manner. It has been suggested
 22 that ROS promote angiogenesis, either directly through stimulation of HIF-1 genes that are
 23 involved in stimulating angiogenesis, such as NOS and NADPH oxidase or through the gener-
 24 ation of active oxidation products, including lipid peroxides. ROS are associated with the
 25 development of several chronic diseases that include atherosclerosis, type 2 diabetes mellitus,
 26 and cancer [43]. Although ROS have damaging effects on tissues, causing cell death at high
 27 concentrations, lesser degrees of oxidative stress may play a positive role during angiogene-

01 sis, or other pathophysiological processes. Angiogenesis induced by oxidative stress involves
02 vascular endothelial growth factor (VEGF) signalling, although VEGF-independent pathways
03 have also been identified [44].

04 The clinical importance of this biological process has become increasingly apparent over the
05 last decade, and angiogenesis now represents a major focus for novel therapeutic approaches
06 to the prevention and treatment of multiple diseases, most notably ischemic cardiovascular
07 disease and cancer [10].

08 **4. Atherosclerosis and plaque angiogenesis**

09 Considering the important contributions of HIF-1 in angiogenesis, it may also be a promising
10 target for treating ischaemic disease [1] and pressure-overload heart failure [45].

11 Atherosclerosis causes clinical disease through the occlusion of the arteries as a result of
12 excessive build-up of plaque within the artery wall resulting from the accumulation of choles-
13 terol, fatty material and extracellular matrix. This causes obstruction in the blood flow to the
14 myocardium (coronary heart disease), brain (ischemic stroke) or lower extremities (peripheral
15 vascular). The most common of these manifestations is coronary heart disease that includes
16 stable angina pectoris and the acute coronary syndromes [46].

17 Coronary heart disease (CHD) is a major cause of mortality globally (1 in every 6 deaths
18 annually). An estimated £2bn per annum is used to treat CHD and its co-morbidities [47].
19 Arterial injury plays a key role in the initiation and progression of CHD [48]. Treatments
20 for CHD range from lifestyle changes and non-invasive medical therapies to pharmacologi-
21 cal therapies and open surgical interventions. Despite the widespread use of drugs such as
22 statins, there remains a significant proportion of individuals for whom response to therapy is
23 sub-optimal, and who develop atherosclerosis [49, 50].

24 Atherosclerosis is a lipoprotein-driven disease affecting medium and large arteries that
25 leads to plaque formation at specific sites of the arterial tree through intimal inflamma-
26 tion, necrosis, fibrosis and calcification. It is a chronic inflammatory process that involves
27 increased oxidative stress, endothelial damage, and smooth muscle cell proliferation and
28 migration. It is associated with several established risk factors, including hypertension,
29 hyperglycaemia, ageing and dyslipidaemia [51]. It is important to control the factors
30 involved in the progression of atherosclerosis because advanced atherosclerotic lesions are
31 prone to rupture, leading to disability or death. Plaque at risk of rupture has been a major
32 focus of research [52]. There is an emerging need for new therapies to stabilize athero-
33 sclerotic lesions. Further understanding of the effects of hypoxia in atherosclerotic lesions
34 could indicate potential therapeutic targets [53, 54]. The presence of hypoxia in human
35 carotid atherosclerotic lesions correlates with angiogenesis. Hypoxia plays a key role in
36 the progression and development of advanced lesions by promoting lipid accumulation,
37 increased inflammation, ATP depletion and angiogenesis. A recent study has convincingly
38 demonstrated the presence of hypoxia in macrophage-rich regions of advanced human
39 carotid atherosclerotic lesions [53].

01 4.1. Evidence for hypoxia within atherosclerotic plaque

02 Hypoxia in atherosclerotic plaques is now widely recognized, because of the use of specific
03 probes in imaging studies [4]. Imaging plaque hypoxia could provide a means of assessing
04 putative culprit lesions that are at risk of rupture, and are consequentially liable to adverse
05 outcomes.

06 Hypoxia has been consistently found in atherosclerotic plaques *in vivo* in humans and animal
07 models using different biomarkers [55]. The immunologically identifiable hypoxia marker,
08 7-(4'-(2-nitroimidazole-1-yl)-butyl)-theophylline (NITP), has been used to assess hypoxia in
09 three murine models *in vivo*. NITP can bind to cells under low-oxygen conditions [56, 57].

10 Other non-invasive imaging techniques have also been applied, which directly target plaque
11 hypoxia, and these techniques are now being further validated in human studies. The met-
12 abolic marker F-fluorodeoxyglucose (FDG) has been used to detect human atherosclerosis
13 *in vivo* and may also serve as an indirect marker of plaque hypoxia as the enhanced glu-
14 cose uptake in anaerobic metabolism results in an increased uptake of the labelled FDG
15 [58]. F-18-fluoromisonidazole positron emission tomographic (PET) has been used for the *in*
16 *in vivo* assessment of hypoxia in advanced aortic atherosclerosis in rabbits where hypoxia has
17 been found to be predominantly confined to the macrophage-rich regions within the ath-
18 eromatous core, whereas the macrophages close to the lumen were hypoxia negative [47].
19 This was then related to hypoxia assessed by *ex vivo* tissue staining using pimonidazole, and
20 immuno-staining for macrophages (RAM-11), new vessels (CD31) and hypoxia-inducible
21 factor-1 α . ^{18}F -fluoromisonidazole (^{18}F -FMISO), a cell permeable 2-nitroimidazole derivative
22 that is reduced *in vivo* by nitroreductases, regardless of the intracellular oxygen concentra-
23 tion, has been one of the leading radiotracers for imaging hypoxia [47]. In human studies,
24 this imaging approach has been coupled with quantitative polymerase chain reaction (qPCR)
25 and immuno-staining of plaques tissues recovered by carotid endarterectomy to determine
26 the gene expression of HIF-1 α and cluster of differentiation 68 (CD68, a marker of inflam-
27 mation). HIF-1 α and CD68 expression were both found to be significantly correlated with
28 F-FDG-uptake, indicating an association between the presence of hypoxia, inflammation and
29 increased glucose metabolism *in vivo* [59].

30 Imaging plaque biomarkers such as CRP, interleukins 6, 10 and 18, soluble CD40 ligand, P-
31 and E-selectin, NT-proBNP, fibrinogen and cystatin C show great potential in the prediction
32 and improvement for vascular patients [60].

33 4.2. The development of a hypoxic environment within the atherosclerotic plaque

34 Hypoxia has been identified as a potential factor in the formation of vulnerable plaque, and
35 it is clear that decreased oxygen plays a role in the development of plaque angiogenesis lead-
36 ing to plaque destabilization [61]. There have been a number of hypotheses of atherogenesis
37 (plaque angiogenesis) proposing that an imbalance between the demand for and supply of
38 oxygen in the arterial wall is a key factor in the development of atherosclerosis [2, 62].

39 During atherogenesis, the intima (the innermost layer of the artery wall) may thicken by the
40 accumulation of cells and matrix, and the diffusion of oxygen can then become impaired. The

01 vasa vasorum, forming the network of small blood vessels, are vulnerable to hypoxia espe-
02 cially at the site of arterial branching as they are end arteries and the blood flow is reduced in
03 this region. It has been hypothesized that hypoxia within the vasa vasorum is due to reduced
04 blood flow and consequent endothelial dysfunction, local inflammation and permeation of
05 large particles such as microbes, LDL-lipoprotein and fatty acids which are transformed by
06 macrophages into foam cells [63, 64], which may be an initiating factor in atherosclerosis
07 [65]. Therefore, the micro-environment within the atherosclerotic plaque is thought to be an
08 important determinant of whether a plaque progresses, and the likelihood of clinical compli-
09 cations. Recent reports provide substantial evidence that there are regions within the plaque
10 in which hypoxia can be identified [66].

11 In addition to being a marker of hypoxia, HIF-1 α may directly enhance atherogenesis through
12 several mechanisms, including smooth muscle cell proliferation and migration, new vessel
13 formation (angiogenesis) and altered lipid metabolism [67]. The effects of HIF-1 α on macro-
14 phage biology and subsequent promotion of atherogenesis has been studied in mice. HIF-1 α
15 expression in macrophages affects their intrinsic inflammatory profile and promotes the
16 development of atherosclerosis [68]. Hence, HIF-1 α may play a key role in the progression
17 of atherosclerosis by initiating and promoting the formation of foam cells, endothelial cell
18 dysfunction, apoptosis, increasing inflammation and angiogenesis [69].

19 It has been also proposed that the state of hypoxia, present in the atherosclerotic plaques of
20 mice deficient in apolipoprotein E (ApoE^{-/-} mice), may promote lipid synthesis, and reduce
21 cholesterol efflux through the ATP-binding cassette transporter (ABCA1) pathway: processes
22 that are known to be mediated by HIF-1 α [55]. Hypoxia has also been reported to increase
23 the formation of lipid droplets in macrophages to promote the secretion of inflammatory
24 mediators, and atherosclerotic lesion progression by exacerbating ATP depletion and lactate
25 accumulation in this model of atherosclerosis [53].

26 Several HIF-responsive genes have been found to be upregulated in atherosclerosis, such as
27 VEGF, endothelin-1 and matrix-metalloproteinase-2 [70]. Hypoxia has the potential to fun-
28 damentally change the function, metabolism and responses of many of the cell types found
29 within the developing atherosclerotic plaque, and this may in turn determine whether the
30 plaque evolves into a stable or unstable phenotype. It is likely that this is mediated through
31 effects on angiogenesis, extracellular matrix elaboration and lipoprotein metabolism. The
32 hypoxic milieu in the atherosclerotic plaque may therefore also have implications for the
33 putative therapeutic interventions for atherosclerosis. However, most *in vitro* studies have
34 been conducted under normoxic conditions. The effects observed under these conditions may
35 not accurately reflect those extant within the plaque [70].

36 The role of HIF-1 in atherosclerosis is not univocal. Silencing of HIF-1 α in macrophages reduces
37 proinflammatory factors and increases macrophage apoptosis. Hyperlipidaemia impairs
38 angiogenesis in an HIF-1 β and nuclear factor (NF)- κ B-dependent manner. Specific knockdown
39 of HIF-1 α in endothelial cells reduces atherosclerosis through reduced monocyte recruitment
40 [26], whereas knockdown in antigen-presenting cells results in aggravation of atherosclerosis
41 through T-cell polarization [71]. There is another non-lipid-driven mechanism by which alter-
42 native macrophages present in human atherosclerosis M(Hb) promote plaque neoangiogenesis
43 and microvessel incompetence through an HIF-1 α /VEGF-A-dependent pathway [72].

01 HIF-1 α has also been also implicated in the pathogenesis of in-stent restenosis following
02 coronary revascularisation, stroke, peripheral artery disease, aortic aneurysm formation and
03 pulmonary artery hypertension [73], and also appears to be involved in the calcification of
04 blood vessels, which often accompanies atherosclerosis [74]. Despite being an intracellular
05 transcription factor, HIF-1 could be possible released into the circulation from damaged cells,
06 similar to other transcriptional factors such as NF- κ B and p53 [74-76].

07 4.3. Other atherogenic mechanisms of hypoxia

08 Although plaque angiogenesis is a physiological response that facilitates the increased oxygen
09 demand in the plaque, it can have adverse effects by facilitating intra-plaque haemorrhage
10 (IPH) and the influx of inflammatory mediators. IPH as a result of immature plaque neovessels
11 is associated with subsequent ischemic events. Inflammatory cell, endothelial cell and pericyte
12 interactions can provide insight into the biological mechanisms of plaque angiogenesis [71].

13 The recruitment of T lymphocytes and proliferation and migration of smooth muscle and
14 endothelial cells are essential for atherosclerotic plaque formation and development. During
15 this process, a number of pro-inflammatory factors and cytokines, leukotrienes and chemo-
16 kines are increased in expression, especially in lipid-loaded foam cells, such as IL8, tumour
17 necrosis factor α (TNF α), interleukin (IL)-1,vascular cell adhesion molecule 1 (VCAM-1) and
18 15-lipoxygenase-2 (15-LOX-2). Moreover, macrophages are trapped in hypoxic areas of the
19 lesion; however, the exact mechanisms have yet to be determined.

20 The majority of inflammatory cells contributing to early atherosclerosis probably enter the
21 artery wall from the lumen [77, 78]. However, the vasa vasorum and associated microvessels
22 may provide an alternate route by which leucocytes can enter the vascular wall [79]. As ath-
23 erosclerosis progresses, angiogenic factors within the micro-environment of the plaque may
24 stimulate new vessel formation. This combination of delicate new vessel network and inflam-
25 matory cells, that elaborate proteolytic enzymes, may contribute to intra-plaque haemorrhage
26 and subsequent plaque rupture [80]. The involvement of vasa vasorum and intimal hyper-
27 plasia in the pathophysiology of atherosclerosis is supported by several experimental animal
28 studies [81, 82].

29 Hypoxia may also induce macrophage migration inhibitory factor (MIF). MIF plays a critical
30 role in the progression of atherosclerosis by several different mechanisms. These include the
31 MIF-triggered arrest and chemotaxis of monocytes and T cells through its receptors CXCR2/4.
32 Further, *in vivo* studies have shown that the blockade of MIF in mice with advanced athero-
33 sclerosis leads to plaque regression and reduced monocyte and T-cell content. Additionally,
34 the neuronal signalling molecule Netrin-1 was recently shown to play an important role
35 in macrophage retention in atherosclerotic plaques. Notably, netrin-1 expression has been
36 shown to be regulated by hypoxia, but this may be tissue or disease specific [55].

37 Atherosclerotic lesion formation is associated with vessel wall thickening resulting in
38 regional limited oxygen exchange. Vascular cells respond to hypoxic conditions with
39 changes in cell metabolism, angiogenesis, apoptosis and inflammatory responses compa-
40 rable to cells in tumours. Local hypoxic regions and hypoxic cells have been identified in
41 human atherosclerotic lesions and in experimental models. Increased oxygen consumption
42 by cells with a high metabolic activity, such as macrophages, further depletes the oxygen

01 availability, creating a hypoxic environment in the atherosclerotic lesion. In macrophages,
02 hypoxia not only affects the metabolism and lipid uptake but also results in an increased
03 inflammatory response characterized by increased IL-1 β and caspase-1 activation. Hypoxia
04 also augments the thrombotic potential of atherosclerotic plaques through upregulation
05 of tissue factor.

06 The identification of specific inflammatory markers pertaining to the arterial wall in ath-
07 erosclerosis may be useful for both diagnosis and treatment. These include macrophage
08 inhibiting factor (MIF), leucocytes and P-selectin. Purinergic signalling is involved in the
09 control of vascular tone and remodelling. Endothelial cells release purines and pyrimi-
10 dines in response to changes in blood flow (evoking shear stress) and hypoxia. They then
11 act on P2Y, P2X and P1 receptors on endothelial cells leading to release of EDRF mediated
12 by nitric oxide and prostaglandins and EDHF, resulting in vasodilatation. The therapeutic
13 potential of purinergic compounds for the treatment of vascular diseases, including hyper-
14 tension, ischaemia, atherosclerosis, migraine and coronary artery and diabetic vascular
15 disease as well as vasospasm is discussed [83]. Modern therapeutic modalities involv-
16 ing endothelial progenitor cells therapy, angiotensin II type-2 (AT2R) and ATP-activated
17 purinergic receptor therapy are notable to mention. Future drugs may be designed to
18 target three signalling mechanisms of AT2R which are (a) activation of protein phosphatases
19 resulting in protein dephosphorylation, (b) activation of bradykinin/nitric oxide/
20 cyclic guanosine 3',5'-monophosphate pathway by vasodilation and (c) stimulation of
21 phospholipase A(2) and release of arachidonic acid. Drugs may also be designed to act on
22 ATP-activated purinergic receptor channel type P2X7 molecules which acts on cardiovas-
23 cular system. Better understanding of the vascular inflammatory processes and the cells
24 involved in the formation of plaques may prove to be beneficial for future diagnosis, clinical
25 treatment and planning innovative novel anti-atherosclerotic drugs [84].

26 Systemic hypoxia that is, for example, associated with obstructive sleep apnoea (OSA)
27 also promotes atherosclerosis. The processes by which it may do this include effects on
28 lipid metabolism and efflux, inflammation, altered macrophage polarization and glucose
29 metabolism [85].

AQ03

30 5. Conclusion

31 Hypoxia is involved in several pathophysiological processes, including embryogenesis,
32 angiogenesis and atherogenesis. HIF-1 appears to be an important mediator controlling cel-
33 lular response to hypoxia. It also appears to be related to atherosclerotic progression and
34 rupture. A better understanding of the mechanism involved in these processes may provide
35 some novel therapeutic approaches to the treatment of cardiovascular disease.

01 **Author details**

02 L. Heikal* and G.A.A. Ferns

03 *Address all correspondence to: l.heikal@bsms.ac.uk

04 Brighton & Sussex Medical School, Brighton, United Kingdom

AQ01

05 **References**

06 [1] Zimna A and Kurpisz M. Hypoxia-inducible factor-1 in physiological and pathophysi-
07 ological angiogenesis: applications and therapies. *BioMed Research International*.
08 2015;2015DOI: 10.1155/2015/549412

09 [2] Bjornheden T, Levin M, Evaldsson M and Wiklund O. Evidence of hypoxic areas within
10 the arterial wall in vivo. *Arteriosclerosis, Thrombosis and Vascular Biology*. 1999;19(4PT).

AQ04

11 [3] Michiels C, Deleener F, Arnould T, Dieu M and Remacle J. Hypoxia stimulates human
12 endothelial cells to release smooth muscle cell mitogens—role of prostaglandins and
13 BFGF. *Experimental Cell Research*. 1994;213(1):43–54. DOI: 10.1006/excr.1994.1171

14 [4] Humar R, Kiefer FN, Berns H, Resink TJ and Battegay EJ. Hypoxia enhances vascular
15 cell proliferation and angiogenesis in vitro via rapamycin (mTOR) -dependent signaling.
16 *The FASEB Journal*. 2002;16(8):771–780. DOI: 10.1096/fj.01-0658com

17 [5] Krock BL, Skuli N and Simon MC. Hypoxia-induced angiogenesis: good and evil. *Genes
18 & Cancer*. 2011;2(12):1117–33. DOI: 10.1177/1947601911423654

19 [6] Chen L, Endler A and Shibasaki F. Hypoxia and angiogenesis: regulation of hypoxia-
20 inducible factors via novel binding factors. *Experimental and Molecular Medicine*.
21 2009;41:849–857.

22 [7] Chan CK and Vanhoutte PM. Hypoxia, vascular smooth muscles and endothelium. *Acta
23 Pharmaceutica Sinica B*. 2013;3(1):1–7. DOI: 10.1016/j.apsb.2012.12.007

24 [8] Singh RB, Mengi SA, Xu Y-J, Arneja AS and Dhalla NS. Pathogenesis of atherosclerosis:
25 a multifactorial process. *Experimental & Clinical Cardiology*. 2002;7(1):40–53.

26 [9] Cines DB, Pollak ES, Buck CA, et al.. Endothelial cells in physiology and in the patho-
27 physiology of vascular disorders. *Blood*. 1998;91(10):3527–3561.

28 [10] Semenza GL. Regulation of hypoxia-induced angiogenesis: a chaperone escorts VEGF
29 to the dance. *The Journal of Clinical Investigation*. 2014;108(1):39–40. DOI: 10.1172/
30 JCI13374

- 01 [11] Goggins BJ, Chaney C, Radford-Smith GL, Horvat JC and Keely S. Hypoxia and integrin-
02 mediated epithelial restitution during mucosal inflammation. *Frontiers in Immunology*.
03 2013;**4**DOI: 10.3389/fimmu.2013.00272
- 04 [12] Ziello JE, Jovin IS and Huang Y. Hypoxia-inducible factor (HIF)-1 regulatory pathway
05 and its potential for therapeutic intervention in malignancy and ischemia. *The Yale*
06 *Journal of Biology and Medicine*. 2007;**80**(2):51–60.
- 07 [13] Ahluwalia A and Tarnawski AS. Critical role of hypoxia sensor HIF-1 alpha in VEGF gene
08 activation. Implications for angiogenesis and tissue injury healing. *Current Medicinal*
09 *Chemistry*. 2012;**19**(1):90–97.
- 10 [14] Fandrey J. Oxygen-dependent and tissue-specific regulation of erythropoietin gene
11 expression. *American Journal of Physiology–Regulatory, Integrative and Comparative*
12 *Physiology*. 2004;**286**(6):R977–R988. DOI: 10.1152/ajpregu.00577.2003
- 13 [15] Heikal L, Ghezzi P, Mengozzi M, Stelmaszczuk B, Feelisch M and Ferns GAA.
14 Erythropoietin and a nonerythropoietic peptide analog promote aortic endothelial cell
15 repair under hypoxic conditions: role of nitric oxide. *Hypoxia*. 2016;**4**:121–133.
- 16 [16] Heikal L, Ghezzi P, Mengozzi M and Ferns G. Low oxygen tension primes aortic endo-
17 thelial cells to the reparative effect of tissue-protective cytokines. *Molecular Medicine*.
18 2015;**21**:709–716. DOI: 10.2119/molmed.2015.00162
- 19 [17] Myllyharju J. Prolyl 4-hydroxylases, master regulators of the hypoxia response. *Acta*
20 *Physiologica*. 2013;**208**(2):148–165. DOI: 10.1111/apha.12096
- 21 [18] Milkiewicz M, Pugh CW and Egginton S. Inhibition of endogenous HIF inactivation
22 induces angiogenesis in ischaemic skeletal muscles of mice. *The Journal of Physiology*.
23 2004;**560**(1):21–26. DOI: 10.1113/jphysiol.2004.069757
- 24 [19] Provenzano R, Besarab A, Sun CH, et al. Oral hypoxia-inducible factor prolyl hydroxy-
25 lase inhibitor roxadustat (FG-4592) for the treatment of anemia in patients with CKD.
26 *Clinical Journal of the American Society of Nephrology*. 2016;**11**(6):982–991. DOI:
27 10.2215/cjn.06890615
- 28 [20] Jain MR, Joharapurkar AA, Pandya V, et al. Pharmacological characterization of ZYAN1,
29 a novel prolyl hydroxylase inhibitor for the treatment of anemia. *Drug Research*.
30 2016;**66**(2):107–112. DOI: 10.1055/s-0035-1554630
- 31 [21] Beuck S, Schaezner W and Thevis M. Hypoxia-inducible factor stabilizers and other
32 small-molecule erythropoiesis-stimulating agents in current and preventive doping
33 analysis. *Drug Testing and Analysis*. 2012;**4**(11):830–845. DOI: 10.1002/dta.390
- 34 [22] Yousaf F and Spinowitz B. Hypoxia-inducible factor stabilizers: a new avenue for reduc-
35 ing BP while helping hemoglobin?. *Current Hypertension Reports*. 2016;**18**(3)DOI:
36 10.1007/s11906-016-0629-6
- 37 [23] Forristal CE, Winkler IG, Nowlan B, Barbier V, Walkinshaw G and Levesque JP..
38 Pharmacologic stabilization of HIF-1 α increases hematopoietic stem cell quiescence in
39 vivo and accelerates blood recovery after severe irradiation. *Blood*. 2013;**121**(5):759–769.
40 DOI: 10.1182/blood-2012-02-408419

- 01 [24] Maxwell PH and Eckardt KU. HIF prolyl hydroxylase inhibitors for the treatment of
02 renal anaemia and beyond. *Nature Reviews in Nephrology*. 2016;**12**(3):157–168. DOI:
03 10.1038/nrneph.2015.193
- 04 [25] Kuschel A, Simon P and Tug S. Functional regulation of HIF-1 α under normoxi-
05 auis there more than post-translational regulation?. *Journal of Cellular Physiology*.
06 2012;**227**(2):514–524. DOI: 10.1002/jcp.22798
- 07 [26] Pajusola K, Kunnapu J, Vuorikoski S, et al. Stabilized HIF-1 α is superior to VEGF
08 for angiogenesis in skeletal muscle via adeno-associated virus gene transfer. *Faseb*
09 *Journal*. 2005;**19**(8):1365. DOI: 10.1096/fj.05-3720fje
- 10 [27] Tsurumi Y, Takeshita S, Chen DF, et al. Direct intramuscular gene transfer of naked
11 DNA encoding vascular endothelial growth factor augments collateral development
12 and tissue perfusion. *Circulation*. 1996;**94**(12):3281–3290.
- 13 [28] Eckle T, Köhler D, Lehmann R, El Kasmī KC and Eltzschig HK. Hypoxia-inducible fac-
14 tor-1 is central to cardioprotection. A New Paradigm for Ischemic Preconditioning.
15 2008;**118**(2):166–175. DOI: 10.1161/circulationaha.107.758516
- 16 [29] Tekin D, Dursun AD and Xi L. Hypoxia inducible factor 1 (HIF-1) and cardioprotection.
17 *Acta Pharmacologica Sinica*. 2010;**31**(9):1085–1094. DOI: 10.1038/aps.2010.132
- 18 [30] Zhao J, Du F, Shen G, Zheng F and Xu B. The role of hypoxia-inducible factor-2 in diges-
19 tive system cancers. *Cell Death Disease*. 2015;**6**:e1600. DOI: 10.1038/cddis.2014.565
- 20 [31] Agnieszka L, Alicja J and Jozef D. HIF-1 and HIF-2 transcription factors - similar but not
21 identical. *Molecules and Cells*. 2010;**29**(5):435–442.
- 22 [32] Carroll VA and Ashcroft M. Role of hypoxia-inducible factor (HIF)-1 α -versus
23 HIF-2 α in the regulation of HIF target genes in response to hypoxia, insulin-like
24 growth factor-1, or loss of von Hippel-Lindau function: implications for targeting
25 the HIF pathway. *Cancer Research*. 2006;**66**(12):6264–6270. DOI: 10.1158/0008-5472.
26 can-05-2519
- 27 [33] Eubank TD, Roda JM, Liu HW, O'Neil T and Marsh CB. Opposing roles for HIF-1 α
28 and HIF-2 α in the regulation of angiogenesis by mononuclear phagocytes. *Blood*.
29 2011;**117**(1):323–332. DOI: 10.1182/blood-2010-01-261792
- 30 [34] Li QF, Wang XR, Yang YW and Lin H. Hypoxia upregulates hypoxia inducible factor
31 (HIF)-3 α expression in lung epithelial cells: characterization and comparison with
32 HIF-1 α . *Cell Research*. 2006;**16**(6):548–558. DOI: 10.1038/sj.cr.7310072
- 33 [35] Pugh CW and Ratcliffe PJ. Regulation of angiogenesis by hypoxia: role of the HIF sys-
34 tem. *Nature Medicine*. 2003;**9**(6):677–684.
- 35 [36] Graham CH, Fitzpatrick TE and McCrae KR. Hypoxia stimulates urokinase receptor
36 expression through a heme protein-dependent pathway. *Blood*. 1998;**91**(9):3300–3307.
- 37 [37] Phelan MW, Forman LW, Perrine SP and Faller DV. Hypoxia increases thrombospon-
38 din-1 transcript and protein in cultured endothelial cells. *Journal of Laboratory and*
39 *Clinical Medicine*. 1998;**132**(6):519–529. DOI: 10.1016/s0022-2143(98)90131-7

- 01 [38] Kuwabara K, Ogawa S, Matsumoto M, et al. Hypoxia-mediated induction of acidic/basic
02 fibroblast growth factor and platelet derived growth factor in mononuclear phagocytes stim-
03 ulates growth of hypoxic endothelial cells. *Proceedings of the National Academy of Sciences*
04 of the United States of America. 1995;**92**(10):4606–4610. DOI: 10.1073/pnas.92.10.4606
- 05 [39] Wykoff CC, Pugh CW, Maxwell PH, Harris AL and Ratcliffe PJ. Identification of
06 novel hypoxia dependent and independent target genes of the von Hippel-Lindau
07 (VHL) tumour suppressor by mRNA differential expression profiling. *Oncogene*.
08 2000;**19**(54):6297–6305. DOI: 10.1038/sj.onc.1204012
- 09 [40] Sakuda H, Nakashima Y, Kuriyama S and Sueishi K. Media conditioned by smooth mus-
10 cle cells cultured in a variety of hypoxic environments stimulates in vitro angiogenesis-
11 a relationship to transforming growth factor-beta-1. *American Journal of Pathology*.
12 1992;**141**(6):1507–1516.
- 13 [41] Phillips PG, Birnby LM and Narendran A. Hypoxia induces capillary network forma-
14 tion in cultured bovine pulmonary microvessel endothelial cells. *American Journal of*
15 *Physiology-Lung Cellular and Molecular Physiology*. 1995;**268**(5):L789–L800.
- 16 [42] Norrby K. Mast cells and angiogenesis. *Apmis*. 2002;**110**(5):355–371. DOI:
17 10.1034/j.1600-0463.2002.100501.x
- 18 [43] Goerlach A, Dimova EY, Petry A, et al. Reactive oxygen species, nutrition, hypoxia
19 and diseases: Problems solved?. *Redox Biology*. 2015;**6**:372–385. DOI: 10.1016/j.
20 redox.2015.08.016
- 21 [44] Kim YW and Byzova TV.. Oxidative stress in angiogenesis and vascular disease. *Blood*.
22 2014;**123**(5):625–631. DOI: 10.1182/blood-2013-09-512749
- 23 [45] Semenza G L. Hypoxia-inducible factor 1 and cardiovascular disease. *Annual Review of*
24 *Physiology*. 2014;**76**:39–56. DOI: 10.1146/annurev-physiol-021113-170322
- 25 [46] Lahoz C and Mostaza JM. Atherosclerosis as a systemic disease. *Revista Espanola De*
26 *Cardiologia*. 2007;**60**(2):184–195. DOI: 10.1157/13099465
- 27 [47] Mateo J, Izquierdo-Garcia D, Badimon JJ, Fayad ZA and Fuster V. Noninvasive assess-
28 ment of hypoxia in rabbit advanced atherosclerosis using (18)F-fluoromisonidazole
29 PET imaging. *Circulation. Cardiovascular imaging*. 2014;**7**(2):312–320. DOI: 10.1161/
30 CIRCIMAGING.113.001084
- 31 [48] Varani J and Ward PA. Mechanisms of endothelial cell injury in acute inflammation.
32 *Shock*. 1994;**2**(5):311–319.
- 33 [49] Andrade PJ, Medeiros MM, Andrade AT, Lima AA. Coronary angioplasty versus CABG:
34 review of randomised trials. *Arquivos Brasileiros De Cardiologia*. 2011;**97**(3):E60–E69.
- 35 [50] Solomon AJ, Gersh BJ. Management of chronic stable angina: medical therapy, percu-
36 taneous transluminal coronary angioplasty and coronary artery bypass graft surgery-
37 Lessons from randomized trials. *Annals of Internal Medicine*. 1998;**128**(3):216–223.

- 01 [51] Xiao W, Jia Z, Zhang Q, Wei C, Wang H and Wu Y. Inflammation and oxidative stress,
02 rather than hypoxia, are predominant factors promoting angiogenesis in the initial
03 phases of atherosclerosis. *Molecular Medical Report*. 2015;**12**(3):3315–3322. DOI:
04 10.3892/mmr.2015.3800
- 05 [52] Nie XY, Randolph GJ, Elvington A, et al. Imaging of hypoxia in mouse atherosclerotic
06 plaques with Cu-64-ATSM. *Nuclear Medicine and Biology*. 2016;**43**(9):534–542. DOI:
07 10.1016/j.nucmedbio.2016.05.011
- 08 [53] Hulten LM and Levin M. The role of hypoxia in atherosclerosis. *Current Opinion in*
09 *Lipidology*. 2009;**20**(5):409–414. DOI: 10.1097/MOL.0b013e3283307be8
- 10 [54] Van der Veken B, De Meyer GRY and Martinet W. Intraplaque neovascularization as a
11 novel therapeutic target in advanced atherosclerosis. *Expert Opinion on Therapeutic*
12 *Targets*. 2016;**20**(10):1247–1257. DOI: 10.1080/14728222.2016.1186650
- 13 [55] Parathath S, Yang Y, Mick S and Fisher EA. Hypoxia in murine atherosclerotic
14 plaques and its adverse effects on macrophages. *Trends in Cardiovascular Medicine*.
15 2013;**23**(3):80–84. DOI: 10.1016/j.tcm.2012.09.004
- 16 [56] Bjornheden T, Evaldsson M and Wiklund O. A method for the assessment of hypoxia
17 in the arterial wall, with potential application in vivo. *Arteriosclerosis, Thrombosis and*
18 *Vascular Biology*. 1996;**16**(1):178–185.
- 19 [57] Webster L, Hodgkiss RJ and Wilson GD. Cell cycle distribution of hypoxia and progres-
20 sion of hypoxic tumour cells in vivo. *British Journal of Cancer*. 1998;**77**(2):227–234.
- 21 [58] Buscombe JR. Exploring the nature of atheroma and cardiovascular inflammation in vivo
22 using positron emission tomography (PET). *British Journal of Radiology*. 2015;**88**(1053)
23 DOI: 10.1259/bjr.20140648
- 24 [59] Pedersen SF, Grabe M, Hag AMF, Hojgaard L, Sillesen H and Kjar A. (18)F-FDG imag-
25 ing of human atherosclerotic carotid plaques reflects gene expression of the key hypoxia
26 marker HIF-1alpha. *American Journal of Nuclear Medicine and Molecular Imaging*.
27 2013;**3**(5):384–92.
- 28 [60] van Lammeren GW, Moll FL, Borst GJD, de Kleijn DPV, de Vries JPPM and Pasterkamp
29 G.. Atherosclerotic plaque biomarkers: beyond the horizon of the vulnerable plaque.
30 *Current Cardiology Reviews*. 2011;**7**(1):22–27. DOI: 10.2174/157340311795677680
- 31 [61] Sluimer JC and Daemen MJ. Novel concepts in atherogenesis: angiogenesis and hypoxia
32 in atherosclerosis. *Journal of Pathology*. 2009;**218**(1):7–29. DOI: 10.1002/path.2518
- 33 [62] Gainer JL. Hypoxia and atherosclerosis. Re-evaluation of the old hypothesis.
34 *Atherosclerosis*. 1987;**68**(3):263–266. DOI: 10.1016/0021-9150(87)90206-1
- 35 [63] Jarvilehto M and Tuohimaa P. Vasa vasorum hypoxia: initiation of atherosclerosis.
36 *Medical Hypotheses*. 2009;**73**(1):40–41. DOI: 10.1016/j.mehy.2008.11.046

- 01 [64] Barger AC, Beeuwkes R, Lainey LL and Silverman KJ. Hypothesis-vasa vasorum and
02 neovascularisation of human coronary arteries- a possible role in the path-physiology
03 of atherosclerosis. *New England Journal of Medicine*. 1984;**310**(3):175–177. DOI: 10.1056/
04 nejm198401193100307
- 05 [65] Ferns G and Heikal L. Atherogenesis and hypoxia. *Angiology*. 2016:1–22. DOI: AQ05
06 10.1177/0003319716662423
- 07 [66] Lim CS, Kiriakidis S, Sandison A, Paleolog EM and Davies AH. Hypoxia-inducible factor
08 pathway and diseases of the vascular wall. *Journal of Vascular Surgery*. 2013;**58**(1):219–
09 230. DOI: 10.1016/j.jvs.2013.02.240
- 10 [67] Aarup A, Pedersen TX, Junker N, et al. Hypoxia-inducible factor-1 α expression in mac-
11 rophages promotes development of atherosclerosis. *Arteriosclerosis, Thrombosis, and*
12 *Vascular Biology*. 2016;DOI: 10.1161/atvbaha.116.307830
- 13 [68] Gao L, Chen Q, Zhou X and Fan L. The role of hypoxia-inducible factor 1 in atherosclerosis.
14 *Journal of Clinical Pathology*. 2012;**65**(10):872–876. DOI: 10.1136/jclinpath-2012-200828
- 15 [69] Sluimer JC, Gasc J-M, van Wanroij JL, et al. Hypoxia, hypoxia-inducible transcription
16 factor, and macrophages in human atherosclerotic plaques are correlated with intra-
17 plaque angiogenesis. *Journal of the American College of Cardiology*. 2008;**51**(13):1258–
18 1265. DOI: 10.1016/j.jacc.2007.12.025
- 19 [70] de Vries MR and Quax PHA. Plaque angiogenesis and its relation to inflammation and ath-
20 erosclerotic plaque destabilisation. *Current Opinion in Lipidology*. 2016;**27**(5):499–506.
- 21 [71] Finn AV, Akahori H, Guo L, et al. Alternative macrophages promote intraplaque angio-
22 genesis and vascular permeability in human atherosclerosis. *Journal of the American*
23 *College of Cardiology*. 2016;**67**(13):2241–2241.
- 24 [72] Kasivisvanathan V, Shalhoub J, Lim CS, Shepherd AC, Thapar A and Davies AH.
25 Hypoxia-inducible factor-1 in arterial disease: a putative therapeutic target. *Current*
26 *Vascular Pharmacology*. 2011;**9**(3):333–349.
- 27 [73] Li G, Lu W-h, Ai R, Yang J-h, Chen F and Tang Z. The relationship between serum
28 hypoxia-inducible factor 1 alpha and coronary artery calcification in asymptomatic type
29 2 diabetic patients. *Cardiovascular Diabetology*. 2014;**13**DOI: 10.1186/1475-2840-13-52
- 30 [74] Ismail S, Mayah W, Battia HE, et al. Plasma nuclear factor kappa B and serum peroxire-
31 doxin 3 in early diagnosis of hepatocellular carcinoma. *Asian Pacific Journal of Cancer*
32 *Prevention: APJCP*. 2015;**16**(4):1657–63.
- 33 [75] Attallah AM, Abdel-Aziz MM, El-Sayed AM and Tabll AA. Detection of serum p53 pro-
34 tein in patients with different gastrointestinal cancers. *Cancer Detection and Prevention*.
35 2003;**27**(2):127–131. DOI: [http://dx.doi.org/10.1016/S0361-090X\(03\)00024-2](http://dx.doi.org/10.1016/S0361-090X(03)00024-2)
- 36 [76] Faggionato A, Ross R and Harker L. Studies of hypercholesterolemia in the non-human pri-
37 mate. 1. Changes that lead to fatty streak formation. *Arteriosclerosis*. 1984;**4**(4):323–340.

- 01 [77] Ross R. Mechanisms of disease - atherosclerosis - an inflammatory disease. *New England*
02 *Journal of Medicine*. 1999;**340**(2):115–126.
- 03 [78] Rademakers T, Douma K, Hackeng TM, et al. Plaque-associated vasa vasorum in aged
04 apolipoprotein E-deficient mice exhibit proatherogenic functional features *In Vivo*.
05 *Arteriosclerosis, Thrombosis and Vascular Biology*. 2013;**33**(2):249–256. DOI: 10.1161/
06 atvbaha.112.300087
- 07 [79] Moreno PR, Purushothaman KR, Zias E, Sanz J and Fuster V. Neovascularization
08 in human atherosclerosis. *Current Molecular Medicine*. 2006;**6**(5):457–477. DOI:
09 10.2174/156652406778018635
- 10 [80] Barker SGE, Talbert A, Cottam S, Baskerville PA and Martin JF. Arterial intimal hyper-
11 plasia after occlusion of the adventitial vasa vasorum in the pig. *Arteriosclerosis and*
12 *Thrombosis*. 1993;**13**(1):70–77.
- 13 [81] Khurana R, Zhuang Z, Bhardwaj S, et al. Angiogenesis-dependent and independent
14 phases of intimal hyperplasia. *Circulation*. 2004;**110**(16):2436–2443. DOI: 10.1161/01.
15 cir.0000145138.25577.f1
- 16 [82] Burnstock G. Purinergic signalling and endothelium. *Current Vascular Pharmacology*.
17 2016;**14**(2):130–145. DOI: 10.2174/1570161114666151202204948
- 18 [83] Thent ZC, Chakraborty C, Mahakkanukrauh P, Kosai N, Rajan R and S D. The molecular AQ06
19 concept of atheromatous plaques. *Current Drug Target*. 2016.
- 20 [84] Marsch E, Sluimer JC and Daemen MJAP. Hypoxia in atherosclerosis and inflammation.
21 *Current Opinion in Lipidology*. 2013;**24**(5):393–400. DOI: 10.1097/MOL.0b013e32836484a4